Synthesis and extraction properties of some lariat ethers derived from the spontaneously resolved guaifenesin, 3-(2-methoxyphenoxy)propane-1,2-diol

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Abstract
Capable of spontaneous resolution rac-3-(2-methoxyphenoxy)propane-1,2-diol, guaifenesin 1 has been proposed as a cheap and readily available enantiopure precursor for the synthesis of nonracemic crown ethers having ligating OAr and OMe arms (lariat ethers). The crowns studied failed to form stable host/guest complexes with amine hydrochloride salts; the effective complexation was achieved using hexafluorophosphate salts. Moderate enantiomeric recognition of R*NH$_2$·HPF$_6$ was achieved with the lariat ethers 11c. As a whole, the enantioselectivity of the extraction is inversely related to the extractive power of the lariat ether.

Keywords: Lariat ethers, guaifenesin, extraction, chiral recognition

Introduction
Cyclic polyethers (crowns) are one of the most popular classes of the synthetic receptors capable of selective binding and transport of organic and inorganic ions and molecules.1 Some nonracemic chiral (usually simply named as “chiral”) crowns show a capacity for chiral recognition and predominantly bind one of the pair of enantiomers.2 The first chiral crown ethers were prepared by Cram et al.3 These enantiopure macrocycles showed from moderate to significant enantiomeric recognition toward biologically interesting ammonium guests.4 By now a large number of different crown ethers have been synthesized and studied for molecular recognition toward free and protonated chiral amino acid esters and/or other amines.5,6

An effective chiral recognition requires that a chiral receptor would be capable of forming sufficiently stable complexes with substrate enantiomers and that a chiral barrier would be present, which reduces the stability of one of the diastereomeric complexes thus formed. It is
agreed that the binding abilities of chiral crown ethers could be enhanced particularly by the presence of a flexible side arm with an electron donor site (the case of so called lariat ethers).

The lariat ethers chiral recognition ability towards amino acid derivatives was recently investigated. The common feature of the studied crowns was the presence in their structure of the chiral OCH₂C*H(CH₂OC₆H₄OMe-p)O unit, the fragment of 3-(4-methoxyphenoxy)propane-1,2-diol. The last compound has been prepared through p-methoxyphenol addition to (S)-glycidol.

In the early work of Gokel et al. it was found that within the pair of 15-crown-5 lariat ethers having (CH₂OC₆H₄OMe) side arms, the ortho-methoxy derivative was much more effective host for sodium cation than the para-methoxy one. The obvious reason for this lies in the fact that two oxygen atoms of the substituent could be involved in the cation binding in the case of ortho, whereas the oxygen atom of the p-methoxyl could not be involved in this complexation by steric reasons. Having these facts in mind we decided that 3-(2-methoxyphenoxy)propane-1,2-diol 1, chiral drug guaifenesin, would be the more suitable starting material for the synthesis of the lariat ethers capable of chiral recognition.

Some years ago we have disclosed the conglomerate–forming nature of guaifenesin and developed cheap and very effective resolution procedure based on this property. In the present investigation we have used both enantiomers of guaifenesin prepared through entrainment procedure for construction of the series of nonracemic lariat crown ethers. In addition to that, the binding and chiral recognition ability of the so obtained hosts with respect to α-phenylethylammonium salts and two α-amino acid methyl ester salts is discussed here.

### Results and Discussion

#### Synthesis

For the synthesis of chiral lariat ethers 9-11 the documented general approaches were used. Scheme 1 is representative of the major synthetic sequences.

Both enantiomers of the key starting diol guaifenesin 1 were obtained through spontaneous resolution of the racemic material according to our published procedure. Both enantiomeric building block diols 3 were prepared by the reaction of diol 1 and tosylate 4, which in turn was obtained via dihydroxyrylanl monoprotection of ethylene glycol.

Macrocycles 9 and 10 have been synthesized in 15-31\% yield by the ring closure of chiral subunit diol 1 with tri- or tetraethylene glycol di(p-toluenesulfonate) in the presence of NaH in THF under high dilution conditions. The 18-membered cyclic ethers 11a,b and the 20-membered cyclic ethers 11c have been synthesized by the reaction of enantiomerically pure diols 1 or 3 with the appropriate ditosylates 6 or 8 in the presence of base. The ditosylates 6a-c were synthesized by the reactions of dihydroxyaromatic compounds, such as catechol 2a, 2,3-dihydroxynaphthalene 2b and 1,1′-bi-2-naphthol (S)-2c with ethylene oxide or chloroethanol followed by the interaction of the obtained diols 5 with p-toluenesulfonylchloride.
Scheme 1. Reagents and conditions: (i) preferential crystallization; (ii) NaH, THF, reflux; (iii) HCl, MeOH; (iv) piperidine hydrochloride; (v) TsCl, Py; (vi) K₂CO₃, DMF, 110 °C; (vii) TsCl, DMAP, Et₃N, CH₂Cl₂, r.t.
The crown ether (R)-11a was prepared by ring closure of diol (R)-3 with ditosylate 6a in 15% yield. The crown ethers (S)-11b and (aS,S)-11c were prepared by ring closure of diol (S)-3 with ditosylate 6b or (aS)-6c in 21% and 17% yield, respectively. The yield of the crowns increased approximately two times if the cyclization of diol and ditosylate 8 was explored. Thus, the crown ethers (R)-11b and (aR,R)-11c were prepared by ring closure of diol (R)-1 with ditosylate 8b or (aR)-8c in 44% and 41% yield, respectively.

Lariat ethers 10, 11b, and 11c were prepared as both enantiomers. Only (R)-enantiomers were obtained in the case of 9 and 11a. Crowns 11c contain two different chirality elements in their structures, the centre and the axis. The last element is the attribute of nonracemic (aR)- or (aS)-BINOL 2c. We have tested only enantiomeric (aR,R)- and (aS,S)-diastereomers.

For the purpose of comparison the extraction effectiveness of the lariat ethers with OMe group in ortho- and para-position in the benzene ring we have prepared also (S)-[(4-methoxyphenoxy)methyl]-15-crown-5, (S)-10a; the compound was synthesized through the above outlined approach starting with (S)-3-(4-methoxyphenoxy)propane-1,2-diol (S)-1a.

\[
\text{(S)-10a}
\]

The structures of the all obtained lariat ethers were consistent with the \(^1\)H NMR, \(^{13}\)C NMR and mass spectra.

**Molecular recognition. \(^1\)H NMR experiments**

We have employed NMR spectroscopy to detect and quantify molecular recognition of lariat ethers 9-11 as host molecules. The set of chiral guests investigated in our paper consists of alanine and phenylglycine methyl esters 12 and 13, and α-phenylethylamine 14.

\[
\text{Scheme 2. Guest molecules studied.}
\]

In attempt to find the evidences of the supramolecular binding between the lariat ethers and amine guests we have investigated CDCl\(_3\) solutions of (R)-10 and hydrochlorides of 12 and 14. It was found that \(^1\)H chemical shifts perturbations (\(^1\)H CSPs) in (R)-10 + 12HCl and (R)-10 + 14HCl systems (for both enantiomers of the amine components) relative to individual host or guest solutions were less than 0.02 ppm, and hence they cannot be used as indicators of complex formation. Furthermore, increasing of concentration (from 1 to 10 mM) of the individual hydrochloride salts in CDCl\(_3\) solutions leads to notable (ca 0.2 ppm) CSPs of CH- and Me...
protons for 12-HCl and to negligible CSPs for 14-HCl. Thus, self-association of 12-HCl molecules couldn’t be excluded.

As a whole it is evident that 1H CSs are not very useful instrument in the case of the hydrochloride salts, which is in complete agreement with early Cram et al. observations. Therefore to get insight of the phenomenon under question, an additional NMR method, namely DOSY, that provides diffusivity information, which is expressed in terms of self-diffusion coefficient D, was invoked. The simple physical meaning of the self-diffusion coefficient (the bigger the D the smaller the molecular system mass and vice versa) make for its wide use in complexation studies.

Before to study binary systems, the self-associative properties of the individual components were analyzed (Table 1). Because of almost no change of the D((R)-10) with concentration it is conceivable that no self-association of the host (R)-10 in solution was occurred. On the contrary, decreasing of D with increasing of concentration proves that 14-HCl and, in particular, 12-HCl self-associates. Therefore self-diffusion coefficients of the guests (12-HCl and 14-HCl) in the binary systems (10 + 12-HCl) and (10 + 14-HCl) can hardly be used to monitor complexation between the components because the salts molecules are in fast exchange between free, self-aggregated and complexed states and its self-diffusion coefficients are weighted average. On the other hand almost no changes of D(10) in the studied systems (Table 1) indicates the absence of the more or less stable complexes formation between the host and these guests. This observation correlates well with the early Cram conclusion.

Table 1. Self-diffusion coefficient D (×10⁻⁹ m²·s⁻¹) of individual crown (R)-10 and alkylammonium hydrochloride salts (12-HCl, 14-HCl) and their combinations, in CDCl₃ at T = 298K. Standard deviations are less than 0.08 × 10⁻⁹ m²·s⁻¹

<table>
<thead>
<tr>
<th>Sample</th>
<th>c, mM</th>
<th>D of crown component</th>
<th>D of amine component</th>
</tr>
</thead>
<tbody>
<tr>
<td>(R)-10 (M=356)</td>
<td>1</td>
<td>1.02</td>
<td></td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>1.05</td>
<td></td>
</tr>
<tr>
<td>12·HCl (M=105)</td>
<td>1</td>
<td></td>
<td>1.80</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td></td>
<td>0.66</td>
</tr>
<tr>
<td>14·HCl (M=122)</td>
<td>1</td>
<td></td>
<td>0.81</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td></td>
<td>0.73</td>
</tr>
<tr>
<td>(R)-10+(S)-12·HCl</td>
<td>10</td>
<td>0.92</td>
<td>0.68</td>
</tr>
<tr>
<td>(R)-10+(R)-12·HCl</td>
<td>10</td>
<td>1.04</td>
<td>0.84</td>
</tr>
<tr>
<td>(R)-10+(S)-14·HCl</td>
<td>10</td>
<td>1.03</td>
<td>0.94</td>
</tr>
<tr>
<td>(R)-10+(R)-14·HCl</td>
<td>10</td>
<td>1.04</td>
<td>0.88</td>
</tr>
</tbody>
</table>

In the cited work Cram et al.⁴ established that a precondition for observing diastereomeric complexes of crown ethers with amine salts was that X⁻ of RNH₃⁺X⁻ be unable to hydrogen bond strongly with NH₃⁺. They found that the PF₆⁻, AsF₆⁻, and SbF₆⁻ ions fulfilled this
condition, whereas F\(^-\), Cl\(^-\), Br\(^-\), SCN\(^-\), and CCl\(_3\)CO\(^-\) did not. Choosing the RNH\(_3\)^+PF\(_6^-\) salts for our subsequent experiments we were guided by these recommendations. However salts with PF\(_6^-\) anion are found to be non-soluble in CDCl\(_3\). Thus D of individual solutions of guests couldn't be determined. For this reason we have used \(^1\)H NMR monitored liquid-liquid extraction for the binding properties evaluation in our concluding experiments.

Hexafluorophosphate salts of the amines 12-14 are not accessible as solids, so they were produced in D\(_2\)O solution by ion exchange with excess of LiPF\(_6\) and the corresponding amine hydrochloride. The host 9-11 solution in CDCl\(_3\) (0.01 M, 1 equiv) was used to extract ~ 3 equiv of aqueous RNH\(_3\)^+PF\(_6^-\) salts. The LiPF\(_6\) not only served as the source of the extractible PF\(_6^-\) anion, but Li\(^+\), Cl\(^-\), and excess PF\(_6^-\) ions “salted out” the organic guest complex from D\(_2\)O layer to CDCl\(_3\) one. After extraction the organic phase was dried and analyzed. Typically, the relative concentrations of the enantiomeric guests in CDCl\(_3\) layer were determined from the \(^1\)H NMR spectra of their diastereomeric complexes. Analysis of intensities of guest’s and host’s signals in the \(^1\)H NMR spectra of studied systems allows estimating the relative quantities of the corresponding substances (Table 2). The \(^1\)H NMR chemical shifts of the CH\(_3\), NC*H protons (chiral carbon is marked by asterisk) of the guests are represented in Table 2. \(^1\)H NMR integral ratios of CH\(_3\) or NC*H protons to those of Ar (Ph) multiplets are listed in Table 2 as guest/host ratio. In the control experiments no traces of amines were detected in the CDCl\(_3\) phase free of the crown hosts.

Before proceeding to analyze the Table 2 data it must be noted that any enantiomer of methyl alaninate hexafluorophosphate 12·HPF\(_6\) could not be traceable after extraction in the any of the studied host containing organic phase. In accordance with the Cram data, this phenomenon was associated with the great hydrophilicity of the alanine moiety. Thus, only enantiomeric \(\alpha\)-phenylethylammonium hexafluorophosphate 14·HPF\(_6\) and methyl phenylglicinate hexafluorophosphate 13·HPF\(_6\) were used for quantitative experiments.

As it follows from Table 2, runs 1-3, amine extraction is negligible in the case of 12-membered crown 9. The small cavity dimension is the obvious reason for the effect.

As the cavity dimension increases, as in the case of crowns 10 and 11a-b, so does the quantity of amine extracted into the chloroform phase increases too. This is evidenced by increase of the G/H values in Table 2. Some important tendencies could be traced from the runs 5-18. Firstly, the enantioselectivity of extraction is approximately inversely related to the quantity of extracted material. Thus maximum extraction potential demonstrates naphthyl containing crown 11b (runs 15-18). At the same time the enantioselectivity of these extractions is close to zero. Secondly, the effectiveness of the lariat ethers 10, having ortho-OMe substituent in the benzene ring, is approximately ten times larger in the amine guest transfer from water to organic phase than the same ability for lariat ethers 10a, with OMe group in para-position.
Table 2. The guest/host ratio (G/H; CDCl₃, r.t.), observed in the extractions of (S)- or (R)-α-phenylethylammonium hexafluorophosphate 14HPF₆ and (S)- or (R)-methyl phenylglicinate hexafluorophosphate 13HPF₆ by crowns 9-11

<table>
<thead>
<tr>
<th>Run</th>
<th>Host</th>
<th>Guest</th>
<th>G/H</th>
<th>CRFᵃ</th>
<th>δ (CH₃)</th>
<th>δ (C*H)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>(R)-9</td>
<td>(S)-13HPF₆</td>
<td>0.0</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>(R)-9</td>
<td>(S)-13HPF₆</td>
<td>0.0</td>
<td>-</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>(S)-14HPF₆</td>
<td>0.0</td>
<td>-</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>(R)-14HPF₆</td>
<td>trᵇ</td>
<td>-</td>
<td>1.67ᵇ</td>
<td>4.59</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>(S)-10</td>
<td>(S)-14HPF₆</td>
<td>0.48</td>
<td>~1.2</td>
<td>1.62ᵇ</td>
<td>4.45</td>
</tr>
<tr>
<td>6</td>
<td>(R)-10</td>
<td>(R)-14HPF₆</td>
<td>0.56</td>
<td>0.8</td>
<td>1.63ᵇ</td>
<td>4.42</td>
</tr>
<tr>
<td>7</td>
<td>(S)-10</td>
<td>(S)-13HPF₆</td>
<td>0.25</td>
<td>~0.8</td>
<td>5.12</td>
<td></td>
</tr>
<tr>
<td>8</td>
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<td>(R)-13HPF₆</td>
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<td>5.05</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>(S)-10a</td>
<td>(S)-14HPF₆</td>
<td>0.06</td>
<td>-</td>
<td>1.69ᵇ</td>
<td>4.49</td>
</tr>
<tr>
<td>10</td>
<td>(R)-10a</td>
<td>(R)-14HPF₆</td>
<td>trᵇ</td>
<td>-</td>
<td>1.69ᵇ</td>
<td>4.49</td>
</tr>
<tr>
<td>11</td>
<td>(R)-11a</td>
<td>(S)-14HPF₆</td>
<td>0.32</td>
<td>0.9</td>
<td>1.51ᵇ</td>
<td></td>
</tr>
<tr>
<td>12</td>
<td>(R)-11a</td>
<td>(R)-14HPF₆</td>
<td>0.29</td>
<td>-</td>
<td>1.51ᵇ</td>
<td></td>
</tr>
<tr>
<td>13</td>
<td>(S)-11a</td>
<td>(S)-13HPF₆</td>
<td>0.39</td>
<td>0.9</td>
<td>3.56</td>
<td>4.86</td>
</tr>
<tr>
<td>14</td>
<td>(R)-11a</td>
<td>(R)-13HPF₆</td>
<td>0.35</td>
<td>1.1</td>
<td>3.52</td>
<td>4.80</td>
</tr>
<tr>
<td>15</td>
<td>(S)-11b</td>
<td>(S)-14HPF₆</td>
<td>0.67</td>
<td>~1</td>
<td>1.47ᵇ</td>
<td>4.31</td>
</tr>
<tr>
<td>16</td>
<td>(R)-11b</td>
<td>(R)-14HPF₆</td>
<td>0.64</td>
<td>1.1</td>
<td>1.49ᵇ</td>
<td>4.32</td>
</tr>
<tr>
<td>17</td>
<td>(S)-11b</td>
<td>(S)-13HPF₆</td>
<td>0.54</td>
<td>1.1</td>
<td>3.48</td>
<td>4.80</td>
</tr>
<tr>
<td>18</td>
<td>(R)-11b</td>
<td>(R)-13HPF₆</td>
<td>0.62</td>
<td>1.1</td>
<td>3.50</td>
<td>4.86</td>
</tr>
<tr>
<td>19</td>
<td>(aS,S)-11c</td>
<td>(S)-14HPF₆</td>
<td>0.22</td>
<td>high</td>
<td>1.44ᵇ</td>
<td>4.33</td>
</tr>
<tr>
<td>20</td>
<td>(aS,S)-11c</td>
<td>(R)-14HPF₆</td>
<td>0.0</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>21</td>
<td>(aS,S)-11c</td>
<td>(S)-13HPF₆</td>
<td>0.07</td>
<td>2.5</td>
<td>4.98</td>
<td></td>
</tr>
<tr>
<td>22</td>
<td>(aS,S)-11c</td>
<td>(R)-13HPF₆</td>
<td>0.17</td>
<td></td>
<td>5.05</td>
<td></td>
</tr>
</tbody>
</table>

ᵃCRF - chiral recognition factor, [G]ᵣ/[G]ₛ, where [G]ᵣ and [G]ₛ is the relative amounts of R- or S-guest extracted into CDCl₃ phase.ᵇd, J = 7 Hz. ᵗ traces.

An introduction of the BINOL chiral moiety instead of catechol fragment in the structure of crown 11a not only enlarges the macrocycle cavity in the case of 11c, but disturbs it in the asymmetric way. As a result, the general complexation ability for the crowns 11c decreases, yet the enantioselectivity of this complexation increases. Thus, during amine salt extraction by crown host (aS,S)-11c about 22% of the (S)-14HPF₆ finds its way into chloroform phase, whereas (R)-14HPF₆ does not (runs 19-20).

As one could see, for the 11c+13HPF₆ system (runs 21-22) there is a noticeable difference in extraction for different enantiomers, too. That’s why this system was used for quantitative extraction of rac-13HPF₆ D₂O solutions with CDCl₃ solutions of optically pure host (aS,S)-11c.
During this experiment the layers were equilibrated, the neutral guest was isolated from the complex transferring into organic layer, and the optical rotations of the individual 13 were taken. The isolated by this means phenylglycine methyl ester has had an optical purity of 22%, enriched in the (R)-enantiomer.

Conclusions

Summering up, we could say that the conglomerate nature and the developed on this basis the effective procedure of direct resolution of guaifenesin 1 enable an easy access to the family of enantiomeric lariat crown ethers 9-11. These crowns show no evidence of host/guest association with hydrochloride salts of chiral amines. In the same time these crowns show a capacity for host/guest binding of some ammonium hexafluorophosphate salts which is embodied in the extraction of the guest molecules from aqueous to organic phase. The effectiveness of the lariat ethers 10, having ortho-OMe substituent in the benzene ring, is larger than the same ability for ethers 10a, with OMe group in para-position. Moderate enantiomeric recognition of organic ammonium hexafluorophosphates was achieved with lariat ethers 11c. By and large, the enantioselectivity of extraction is approximately inversely related to the quantity of extracted material.

Experimental Section

General. The NMR spectra were recorded on a Bruker Avance-600 spectrometer (600.13 MHz for $^1$H; 150.864 MHz for $^{13}$C) in CDCl$_3$ with TMS or the signals of the solvent as the internal standard. Optical rotations were measured on a Perkin-Elmer model 341 polarimeter (concentration c is given as g/100 mL). Melting points were determined using a Boëtius apparatus and are uncorrected. The UV-Vis spectra were measured by a Perkin Elmer Lambda 35UV spectrometer. HPLC analyses were performed on a Shimadzu LC-20AD system controller, and UV monitor 275 nm was used as a detector. As a rule, the column used, from Daicel, Inc., was Chiralcel OD (0.46 x 25 cm); column temperature 40 °C; flow rate: 0.4 ml/min. Mass spectra EI were recorded on a mass-spectrometer MAT-212, mass spectra MALDI-TOF were recorded on a mass-spectrometer ULTRAFLEx III.

Synthesis

4-Dimethylaminopyridine, 2-(2-chloroethoxy)ethanol 2,3-dihydroxynaphtalene, lithium hexafluorophosphate, D-alanine methyl ester hydrochloride were purchased from Alfa Aesar. L-Alanine methyl ester hydrochloride was purchased from Fluka. D-Alanine methyl ester hydrochloride was purchased from Aldrich. (R)- and (S)-α-phenyl ethylamine, D(-) and L(+) alpha-phenylglycine, were purchased from Acros Organics. Racemic, D- and L-methyl
phenylglycinate hydrochloride were prepared according to literature procedure.4 The hydrochloride salts of (R)- and (S)-α-phenyl ethylamine were prepared by bubbling HCl gas into solutions of the amine in dry ether. All reactions were carried out in a dry argon atmosphere. All organic solutions and reagents were purified and dried, following standard procedures. Racemic guaifenesin, 3-(2-methoxyphenoxy)propane-1,2-diol, rac-1 is commercially available (Alfa Aesar, A16827); (R)- and (S)-3-(2-methoxyphenoxy)-propane-1,2-diol were prepared from racemate by entrainment resolution method according to previously described protocol.10

Resolution of racemic 3-(2-methoxyphenoxy)-1,2-propanediol (Guaifenesin, rac-1) by preferential crystallization (entrainment)

Racemic guaifenesin rac-1 (90.46 g) and (R)-1 (9.54 g) was dissolved in 800 ml of water at 42-45 °C. The solution was cooled to 23 °C and seeded with finely pulverized (R)-1 (0.25 g). After stirring the mixture for 100 min at 22 ± 0.5 °C, precipitated (R)-1 was collected by filtration (21.26 g, 91 % ee). The extra portion of rac-1 (21.01 g) was then dissolved in the mother liquor at 42 °C; the resulting solution was cooled to 23 °C. After the addition of (S)-1 (0.25 g) as seed crystals to the solution, and stirring the mixture for 110 min at 22 ± 0.5 °C, (S)-1 (19.11 g, 99 % ee) was collected by filtration. Further resolution was carried out at 22 ± 0.5 °C by adding amended amounts of rac-1 to the filtrate in a manner similar to that described above. After second cycle, 17.53 g of (R)-1 (> 99 % ee) and 17.22 g of (S)-1 (84 % ee) were collected. After third cycle, 19.68 g of (R)-1 (87 % ee) and 22.60 g of (S)-1 (89 % ee) were collected. A high degree of enantiomeric purity of collected diols can be achieved by simple recrystallization.

(S)-3-(2-Methoxyphenoxy)propane-1,2-diol, (S)-1. Mp 97-99 °C; [α]D20 = + 9.5 (c 1.0, MeOH); 99.9 % ee (chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; flow rate 1.0 mL/min; tR = 17.4 min).

(R)-3-(2-Methoxyphenoxy)propane-1,2-diol, (R)-1. Mp 97-99 °C; [α]D20 = - 9.4 (c 1.0, MeOH); 99.5 % ee (chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; flow rate 1.0 mL/min; tR = 9.9 min).

(S)-3-(4-Methoxyphenoxy)propane-1,2-diol, (S)-1a was prepared by analogy with a published procedure16 from 4-methoxyphenol and (S)-3-chloropropane-1,2-diol. Mp 79-81°C (n-hexane/benzene) (lit.17 mp 80-82 °C; lit.18 mp 80 °C); [α]D20 = +7.5 (c 0.6, EtOH); 95 % ee [chiral HPLC analysis; Daicel Chiralpak AD-RH (0.46 x 15 cm) column; column temperature 27 °C; eluent: isopropanol/water = 1:3; tR = 18.5 min (minor), tR = 21.8 min (major)]; {lit.17 [α]D20 = +7.3 (c 1.28, EtOH), 88.7 % ee; lit.19 [α]D20 = +7.9 (c 1.0, EtOH), 96% ee}.

1-(Tetrahydropyranoxy)-2-[(p-tolylsulfonyl)-oxy]ethane (4) was prepared by analogy with a published method12 from tosyl chloride (46.0 g, 0.24 mol) and 2-(tetrahydropyranoxy)ethanol (35.28 g, 0.24 mol). Yield 71.0 g (98 %).

(S)-4-(2-Methoxyphenoxy)methyl-3,6-dioxa-1,8-octanediol, (S)-3 was prepared by analogy with a published method.12 To a suspension of NaH (1.68 g, 70 mmol) in 20 mL of THF was added a solution of (S)-3-(2-methoxyphenoxy)propane-1,2-diol, (S)-1 (4.95 g, 25 mmol) in 75 mL of THF at 0 °C. The reaction mixture was stirred for 15 min at 0 °C and then 1 h at room
temperature and refluxed for 2 h. Then a solution of 1-(tetrahydro-pyran-2-yl)sulfanyl)oxy]ethane 4 (15.0 g, 50 mmol) in 70 mL of THF was slowly dropwise added to the refluxing solution for 3 h. When the addition was completed, refluxing was continued for 50 h. After addition of water (300 mL), the mixture was extracted with Et2O (3 x 150 mL). The combined ether layers were washed with brine, dried on MgSO4 and concentrated under reduced pressure. The crude product was dissolved in 45 mL of methanol and acidified with 1 M HCl before pH 2. The reaction mixture was stirred for 8 h and basified with NaHCO3. The water layer was extracted with CH2Cl2 (3 x 150 mL). The combined organic layers were dried on MgSO4 and concentrated under reduced pressure. The crude product was purified by column chromatography (silica gel, EtOAc/light petroleum/EtOH = 14:7:2) to give 1.93 g (27 %) of pure (S)-3 as pale yellow oil; [α]D20 = -2.5 (c 2.0, CHCl3); 99.0 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol = 9:1; flow rate: 1 ml/min; tR = 23.2 min (minor), tR = 28.3 min (major)]. 1H NMR δ: 2.48 (br.s, 1H, OH), 3.14 (br.s, 1H, OH), 3.61-3.63 (m, 2H), 3.67 (dd, J = 10.5, 5.9 Hz, 1H), 3.71-3.79 (m, 7H), 3.84 (s, OCH3), 3.99-4.02 (m, 1H, CH), 4.06 (dd, J = 9.9, 6.6 Hz, 1H), 4.10 (dd, J = 9.9, 4.6 Hz, 1H), 6.88-6.94 (m, 4H, Ar). EI mass spectrum [m/z(%)] C14H22O6: 287(10), 286(53), 149(17), 124(100), 122(7), 109(11), 101(37), 87(21), 77(9), 75(18), 69(6), 61(11), 59(6), 57(26), 45(79), 43(13), 41(11), 31(11).

(R)-4-(2-Methoxyphenoxy)methyl-3,6-dioxo-1,8-octanediol, (R)-3 was synthesized analogously from the (R)-3-(2-methoxyphenoxy)propane-1,2-diol, (R)-1; [α]D20 = +3.0 (c 1.1, CHCl3), [α]D20 = -2.5 (c 1.7, MeOH); 99.2 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol = 9:1; flow rate: 1 ml/min; tR = 24.1 min (major), (tR = 29.6 min (minor)).

2,2'-Dihydroxy-1,1'-binaphthyl, 2c. Racemic 2c was synthesized by a known procedure20 by oxidation of β-naphthol (30.0 g, 0.2 mol) with ferric chloride (56.26 g, 0.2 mol). The yield was 28.6 g (88%). Resolution of rac-2c was carried out, as described previously21 based on different solubility of its complexes with (R,R)-1,2-diaminocyclohexane in toluene; (R)-2c: [α]D20 = + 34.3 (c 1.0, THF); mp 208-210 °C, [lit.22 [α]D20 = + 34.3 (c 1.0, THF); mp 207.5–208.5 °C]. (S)-2c: [α]D20 = -33.8 (c 1.0, THF); mp 208-210 °C, [lit.22 [α]D20 = -33.3 (c 1.0, THF); mp 207-208 °C).

1,2-Bis(2-hydroxyethoxy)benzene (5a) was prepared according to the known procedure8a from catechol (11 g, 100 mmol), diethyamine hydrochloride (as a catalyst) and ethylene oxide (9.8 ml, 200 mmol). The yield was 17.9 g, 91 %; mp 81-83 °C (EtOH/ether 1:1) (lit.8a mp 81-83 °C). 2,3-Bis-(2-hydroxyethoxy)naphthalene (5b) was synthesized according to above described method by using 2,3-dihydroxynaphthalene 2b (1.6 g, 10 mmol), piperidine hydrochloride (as catalyst) and ethylene oxide (0.98 mL, 20 mmol). The yield was 2.21 g (89 %); mp 146-147 °C (lit.8a mp 146-147 °C).

(S)-2,2'-Bis(2-hydroxyethoxy)-1,1'-binaphthyl, (aS)-5c was prepared according to literature procedure13 from (aS)-2c (8.62 g, 30.1 mmol), 2-chloroethanol (8.0 mL, 119 mmol) and K2CO3 (16.62 g, 120 mmol) to give 10.21 g (91 %) of (aS)-5c; mp 130-134 °C; [α]D20 = +26.4 (c 0.8, THF); [lit.13 mp 131-134 °C; [α]S78 = +25.1 (c 0.958, THF)].
(R)-2,2′-Bis(2-hydroxyethoxy)-1,1′-binaphthyl, (aR)-5c was prepared from (aR)-2c in a manner similar to that described for the preparation of (aS)-5c; mp 131-134 °C; [α]D20 = -26.4 (c 0.8, THF); [lit.13 mp 130-134 °C; [α]57820 = -26.4 (c 0.76, THF)].

1,2-Bis-[2-(p-tolylsulfonyl)ethoxy]benzene (6a) was synthesized according to the procedure8a from 1,2-bis-(2-hydroxyethoxy)benzene 5a (9 g, 45.5 mmol), pyridine (40 mL) and p-toluenesulfonylchloride (17 g, 89.3 mmol) at -10 °C to give 21.9 g (96%); mp 95-96 °C (MeOH); [lit.8a mp 95-95.5 °C]. 1H NMR δ: 2.45 (s, 6H, CH3), 4.17 (t, J = 4.6 Hz, 4H), 4.34 (t, J = 4.6 Hz, 4H), 6.82 (dd, J = 5.9, 3.4 Hz, 2H), 6.91 (dd, J = 5.9, 3.3 Hz, 2H), 7.34 (d, J = 8.1 Hz, 4H), 7.81 (d, J = 8.1 Hz, 4H).

2,3-Bis-[2-(p-tolylsulfonyl)ethoxy]naphthalene (6b) was synthesized according to the procedure8a from 5b (2.20 g, 8.9 mmol), p-toluenesulfonylchloride (3.38 g, 17.7 mmol) and pyridine (16 mL) to give 3.92 g (81%); mp 111-112 °C [lit.8a mp 111-112 °C]. 1H NMR δ: 2.41 (s, 6H, CH3), 4.28 (dd, J = 5.2, 4.3 Hz, 4H), 4.43 (dd, J = 5.2, 4.3 Hz, 4H), 7.06 (s, 2H), 7.31 (d, J = 8.1 Hz, 4H), 7.36 (dd, J = 6.2, 2.9 Hz, 2H), 7.63 (dd, J = 6.2, 3.3 Hz, 2H), 7.82 (d, J = 8.1 Hz, 4H).

(S)-2,2′-Bis(2-(p-tolylsulfonyl)ethoxy)-1,1′-binaphthyl, (aS)-6c was prepared by analogy with a published method.13 To a solution of (aS)-5c (5.10 g, 13.6 mmol), triethylamine (4.1 mL, 30 mmol) and 4-(N,N-dimethylamino)pyridine (DMAP) (55 mg, 0.87 mmol) in 300 mL of CH2Cl2 was added p-toluenesulfonylchloride (5.68 g, 29.8 mmol) at 0 °C for 15 min. The reaction mixture was stirred at room temperature for 20 h and subsequently washed with water (5 x 60 mL). The organic layer was dried on MgSO4 and concentrated under reduced pressure to give 8.93 g (96%) of (aS)-6c as white crystals; mp 148-150 °C (toluene/n-hexane = 8:2); [α]D20 = +80.4 (c 1.0, CH2Cl2); [lit.23 mp 144-146 °C; [α]D20 = +80.4 (c 1, CH2Cl2)]. 1H NMR δ: 2.37 (s, 6H), 3.92-3.96 (m, 2H), 4.16-4.19 (m, 2H), 4.26-4.47 (m, 2H), 7.04 (d, J = 8.1 Hz, 4H), 7.11 (d, J = 8.4 Hz, 2H), 7.24 (dd, J = 8.1, 7.1 Hz, 2H), 7.39 (m, J = 8.4, 2.6, 3.7 Hz, 8H), 7.71 (d, J = 8.1 Hz, 2H), 7.99 (d, J = 9.1 Hz, 2H).

(R)-2,2′-Bis(2-(p-tolylsulfonyl)ethoxy)-1,1′-binaphthyl, (aR)-6c was prepared from (aR)-5c in a manner similar to that described for the preparation of (aS)-6c; mp 148-150 °C (toluene/n-hexane = 8:2); [α]D20 = -80.3 (c 1.0, CH2Cl2).

(R)-2,2′-Bis(5-hydroxy-3-oxa-1-pentiloxy)-1,1′-binaphthyl, (aR)-7c. The compound was prepared by analogy with a published method.13 A mixture of (aR)-2c (9.70 g, 33.9 mmol), 2-(2-chloroethoxy)ethanol (16.82 g, 14.2 mL, 135 mmol) and K2CO3 (19.0 g, 137 mmol) in 170 mL of DMF was stirred at 110 °C for 20 h. The reaction mixture was filtrated and concentrated under reduced pressure. The residue was taken up in CH2Cl2 (150 mL) and washed with water (5 x 60 mL) and 2M NaOH (100 mL). The organic layer was dried on MgSO4 and concentrated under reduced pressure to give a viscous oil. The crude product was purified by column chromatography (silica gel, EtOAc/n-hexane = 3:6 and 9:1) to give (aR)-7c (14.42 g, 92%) as a yellow oil; [α]D20 = +29.1 (c 1.5, THF). 1H NMR δ: 3.15-3.21 (m, 4H), 3.37-3.44 (m, 6H), 3.49-3.52 (m, 2H), 4.0-4.03 (m, 2H), 4.14-4.17 (m, 2H), 7.16 (d, J = 11.6 Hz, 2H), 7.22 (dd, J = 8.1, 8.5, 2H), 7.34 (dd, J = 8.1, 7.7 Hz, 2H), 7.44 (d, J = 9.0 Hz, 2H), 7.88 (d, J = 8.1 Hz, 2H), 7.96
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2,3-Bis(5-tosloyloxy-3-oxa-1-pentyloxy)naphthalene (8b) was synthesized as described below for 8c by using 7b^{24} (3.64 g, 9.35 mmol, mp 55-56 °C), Et$_3$N (2.08 g, 20.6 mmol), DMAP (37.4 mg, 0.3 mmol) and p-toluenesulfonylchloride (3.92 g, 20.6 mmol) to give 5.26 g (75.4 %, viscous oil).

(R)-2,2′-Bis(5-tosloyloxy-3-oxa-1-pentyloxy)-1,1′-binaphthyl, (aR)-8c was prepared by analogy with a published method.13 To a solution of (aR)-7c (13.42 g, 29 mmol), Et$_3$N (8.7 mL, 63.8 mmol) and DMAP (0.145 g, 1.16 mmol) in CH$_2$Cl$_2$ (300 mL) was added p-toluenesulfonylchloride (12.29 g, 64.16 mmol) at 0 °C for 15 min. The reaction mixture was stirred at room temperature for 20 h and subsequently washed with water (5 x 60 mL). The organic layer was dried on MgSO$_4$ and concentrated under reduced pressure. The crude product was purified by column chromatography (silica gel, EtOAc/n-hexane = 3:6 and 9:1) that obtained (aR)-8c (20.79 g, 93%) as an orange viscous oil; [α]$_D$^{20} = +31.1 (c 1.0, THF); [lit.25 [α]$_{378}^{25}$ = +31.0 (c 1.0, THF)]. ¹H NMR δ: 2.43 (s, 6H), 2.95-2.98 (m, 2H), 3.01-3.05 (m, 2H), 3.38 (t, J = 4.7 Hz, 4H), 3.68 (t, J = 4.7 Hz, 4H), 3.98-4.06 (m, 4H), 7.14 (d, J = 8.2 Hz, 2H), 7.20 (dd, J = 8.2, 7.0 Hz, 2H), 7.27-7.32 (m, 6H), 7.37 (d, J = 8.8 Hz, 2H), 7.72 (d, J = 8.2 Hz, 4H), 7.83 (d, J = 8.2 Hz, 2H), 7.91 (d, J = 8.9 Hz, 2H). MALDI mass-spectra C$_{42}$H$_{42}$O$_{10}$S$_2$: [M]^+ 770.36, [MH]^+ 771.35, [M + Na]^+ 793.38, [M + K]^+ 809.36.

(R)-[2-Methoxyphenoxy)methyl]-12-crown-4, (R)-9. General procedure for synthesis of crown–ethers

To a suspension of NaH (0.792 g, 33 mmol) in dry THF (100 mL) at 10 °C was added a solution of (R)-1 (1.30 g, 6.6 mmol) in THF (200 mL) for over a period 1.5 h. The reaction mixture was refluxed for 2.5 h. After cooling to 25 °C, a solution of triethylene glycol di-(p-toluenesulphonate) (3.01 g, 6.6 mmol, 1 eq.) in THF (250 mL) was added to reaction mixture over a period of 3 h. The suspension was refluxed for 70 h. The reaction mixture was filtrated and concentrated under reduced pressure. The residue was taken up in CH$_2$Cl$_2$ (150 mL) and subsequently washed with water (3 x 100 mL). The organic layer was dried on MgSO$_4$ and concentrated under reduced pressure to give yellow oil. The crude product was purified by column chromatography (silica gel, EtOAc/n-hexane = 1:1) to give (R)-9 (0.31 g, 15%) as a pale yellow oil; [α]$_D$^{20} = +8.8 (c 5, CHCl$_3$). ¹H NMR δ: 3.57-3.73 (m, 12H, CH$_2$), 3.80-3.87 (m, 5H, CH$_2$, OCH$_3$), 3.90-4.17 (m, 3H, CH, CH$_2$), 6.85-6.91 (m, 4H, Ar); MALDI mass-spectra C$_{16}$H$_{24}$O$_6$: [M]^+ 312.4, [MH]^+ 313.4, [M + Na]^+ 335.4, [M + K]^+ 351.5.

(S)-[2-Methoxyphenoxy)methyl]-15-crown-5, (S)-10 was prepared followed the general procedure from (S)-1 (1.58 g, 7.9 mmol) and tetraethylene glycol di(p-toluenesulphonate) (4.1 g, 7.9 mmol) to yield a pale yellow oil (0.87g, 31%); [α]$_D$^{20} = - 13.1 (c 1.1, CHCl$_3$); 99.1 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/ Et$_2$NH = 80:20:0.1; $r_R$ = 27.1 min (minor), $r_R$ = 41.0 min (major)]; ¹H NMR δ: 3.57-3.82 (m, 21H, CH$_2$, OCH$_3$), 3.90-4.08 (m, 3H, CH, CH$_2$), 6.78-6.89 (m, 4H, Ar). ¹³C NMR δ: 57.5 (OCH$_3$); 70.0, 70.7, 70.81, 70.83, 70.85,
71.06, 71.13, 71.3, 71.5 (CH2); 78.3 (CH), 112.7 (C3Ar), 114.6 (C6Ar), 121.2 (C4Ar), 121.7 (C5Ar), 148.9 (C1Ar), 150.1 (C2Ar). MALDI mass-spectra C18H26O7: [M]+ 356.6, [M+ Na]+ 379.6.

(R)-[2-Methoxyphenoxy)methyl]-15-crown-5, (R)-10. It was prepared from (R)-1 as described for (S)-10; [α]D20 = +13.9 (c 1, CHCl3); 99.8 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; tR = 41.5 min (minor), tR = 26.5 min (major)]. NMR spectra were identical with those cited for (S)-10.

(S)-[(4-Methoxyphenoxy)methyl]-15-crown-5, (S)-10a. The compound was prepared according to a published method8th from (S)-3-(4-methoxyphenoxy)propane-1,2-diol (S)-1a (1.3 g, 6.56 mmol) and tetraethyleneglycol di(p-toluenesulfonate) (3.34 g, 6.6 mmol) to give 0.93 g (40%) of pure product as a pale yellow oil; [α]D20 = -13.8 (c 6.9, CHCl3); [lit.8a [α]D20 = -14.5 (c 7, CHCl3)). 1H NMR δ: 3.65-3.75 (m, 16H, CH2), 3.77 (s, 3H, OCH3), 3.79-3.91 (m, 5H, CH, CH2), 6.81-6.87 (4H, Ar).

(R)-12-[2-Methoxyphenoxy)methyl]-2,3-benzo-18-crown-6, (R)-11a was prepared in a manner similar to that described for the preparation of (R)-9 by using diol (R)-3 (1.3 g, 4.5 mmol), suspension of NaH (0.79 g, 19.9 mmol, 60% in mineral oil) and 1,2-bis-[2-(p-tolylsulfonyl)ethoxy]benzene8a 6a (2.3 g, 4.5 mmol) to yield 0.3 g (15%); [α]D20 = +5.7 (c 1.8, CHCl3). 1H NMR δ: 3.70-3.95 (m, 16H, 4.08-4.22 (m, 8H), 6.88-6.92 (m, 8H, Ar). MALDI mass-spectra C24H32O6: [M]+ 447.93, [M+ Na]+ 470.96, [M+ K]+ 486.94.

(S)-12-[2-Methoxyphenoxy)methyl]-2,3-naphtho-18-crown-6, (S)-11b was prepared in a manner similar to that described for the preparation of (R)-9 by using diol (S)-3 (1.14 g, 4 mmol), suspension of NaH (0.71 g, 17.7 mmol, 60% in mineral oil) and 2,3-bis-[2-(p-tolylsulfonyl)ethoxy]naphthalene8a 6b (2.22 g, 4 mmol). The crude product as a viscous oil was purified by silica gel column chromatography (eluent: hexane/EtOAc from 96:4 until 20:80) to yield 0.42 g (21.5 %). Mp 88-90 °C (EtOH); [α]D20 = -6.0 (c 1.5, CHCl3). 95 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; tR = 41.7 min (major), tR = 47.1 min (minor)]. 1H NMR δ: 3.70-3.82 (m, 12H), 3.91-4.07 (m, 8H), 4.22-4.30 (m, 4H), 6.80-6.87 (m, 4H), 7.10 (d, J = 4.0 Hz, 2H), 7.31 (dd, J = 3.1, 5.8 Hz, 2H), 7.64 (dd, J = 3.6, 5.8 Hz, 2H). 13C NMR δ: 56.0 (CH3O); 68.87, 68.94, 69.31, 69.45, 69.61, 70.40, 71.02, 71.14, 71.29, 71.70 (CH2); 78.03 (CH); 108.02, 108.22, 110.0, 112.33, 114.27, 120.94, 121.45, 124.14, 126.30, 126.36, 129.35, 129.37, 148.57, 149.21, 149.22, 149.78. El mass-spectrum [m/z(1%)] C28H34O8: 499(12), 498(43), 231(16), 213(13), 199(12), 188(8), 187(67), 186(71), 185(10), 171(41), 163(31), 160(9), 159(11), 145(9), 141(6), 131(12), 122(7), 115(11), 102(11), 101(53), 89(16), 87(16), 77(11), 73(11), 59(19), 57(24), 45(100), 43(12), 41(17), 31(13). MALDI mass-spectrum [M]+ 498.3, [MH]+ 499.3, [M+ Na]+ 521.3, [M+ K]+ 537.3.

(R)-12-[2-Methoxyphenoxy)methyl]-2,3-naphtho-18-crown-6, (R)-11b was prepared by analogy with a general procedure, from 1.62 g (8.17 mmol) of (R)-1, suspension of 1.45 g (36.12 mmol, 60% in mineral oil) of NaH and 5.26 g (8.17 mmol) of 8b. The crude product (5.75 g) was purified by silica gel column chromatography (eluent: light petroleum/EtOAc from 10:1 until 1:10). The yield was 1.8 g (44.2 %); Mp 88-90 °C (EtOH); [α]D20 = +5.5 (c 0.9, CHCl3); 94 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; tR = 42.4 min
(major), \( t_R = 47.7 \) min (minor)]. NMR spectra were identical with those cited above for (S)-11b. MALDI mass-spectrum [M]+ 498.3, [MH]+ 499.3, [M + Na]+ 521.3, [M + K]+ 537.3.

(aS,S)-14-[2-Methoxyphenoxy]methyl]-2,3:4,5-di(1,2-naphtho)-20-crown-6, (aS,S)-11c. Method A. (aS,S)-11c was prepared by analogy with the general procedure, from (S)-3 (1.50 g, 5.25 mmol) and (aS)-6c (3.40 g, 4.99 mmol) to give 0.56 g (17%) of pure product as a pale yellow oil; [\( \alpha \)]D20 = -73.1 (c 1.0, THF); 98.5 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; \( t_R = 43.0 \) min (minor), \( t_R = 56.1 \) min (major)]. 1H NMR \( \delta \): 3.37-3.81 (m, 20H, OCH2), 3.97 (s, OMe), 4.15-4.25 (m, 2H), 6.80-6.90 (m, 4H), 6.95 (d, J = 8.6 Hz, 2 H), 7.21 (dd, J = 8.1, 8.7 Hz, 2 H), 7.31 – 7.35 (m, 2H), 7.45 (d, J = 8.5 Hz, 1 H), 7.49 (d, J = 9.0 Hz, 1 H), 7.86 (d, J = 9.1 Hz, 2 H), 7.94 (d, J = 9.0 Hz, 2 H). 13C NMR \( \delta \): 56.0 (CH2O); 69.33, 69.80, 69.88, 70.07, 70.15, 70.20, 70.28, 70.41, 70.46, 70.49, 71.04, 71.09, 71.13, 71.50 (CH2); 77.63 (CH); 112.35, 114.28, 115.85, 116.87, 120.47, 120.97, 121.52, 123.50, 123.60, 123.68, 125.42, 125.50, 126.16, 126.22, 127.81, 127.85, 129.17, 129.19, 129.30, 129.42, 129.58, 129.67, 134.13, 148.56, 149.81, 154.44, 154.71. EI mass-spectrum C30H20O8 \([m/z(\%)]\): 625(51), 624(100), 313(29), 312(16), 199(23), 181(7), 163(18), 158(6), 157(24), 155(21), 145(10), 141(10), 135(8), 121(8), 113(8), 101(34), 92(6), 91(27), 87(10), 83(6), 73(11), 65(14), 59(11), 57(30), 45(79), 43(12), 41(18), 31(15). MALDI mass-spectrum [M]+ 624.4, [MH]+ 625.4, [M + Na]+ 647.4, [M + K]+ 663.5.

(aR,R)-14-[2-Methoxyphenoxy]methyl]-2,3:4,5-di(1,2-naphtho)-20-crown-6, (aR,R)-11c was prepared in a manner similar to that described for the preparation of (aS,S)-11c (method A); [\( \alpha \)]D20 = +74.2 (c 1.0, THF), 99.1 % ee [chiral HPLC analysis: Chiralcel OD; n-hexane/isopropanol/Et2NH = 80:20:0.1; \( t_R = 46.9 \) min (major), \( t_R = 54.6 \) min (minor)]. NMR spectra were identical with those cited above for (aS,S)-11c.

Method B. (aR,R)-11c was prepared by analogy with the general procedure, from (R)-1 (1.50 g, 7.57 mmol) and (aR)-8c (3.54 g, 6.93 mmol) to yield 1.77 g (41%) of a pale yellow oil; [\( \alpha \)]D20 = +74.1 (c 1.0, THF). 1H NMR spectrum was identical with those cited above.

NMR experiments

Self-diffusion coefficients determination. The 2D DOSY experiments were performed by STE-BPLED sequence.26 Data was acquired with a 20-50 ms diffusion delay in all experiments, bipolar gradient pulses duration from 2.3 to 3.5 ms (depending on a system under investigation), 1 ms spoil gradient pulse (30%) and a 5 ms eddy current delay. The bipolar pulse gradient strength was varied incrementally from 0.01 to 0.32 T/m. The experimentally observed diffusion coefficients were then determined from 2D DOSY plots obtained by Bruker XWinNmr software package. Several measures of D were obtained at more than one place in the spectrum and all experiments were carried out in duplicate or triplicate mode. The reported results are the mean value of multiple data points and the standard deviations are less than 0.05·10-9 m2/s in all cases. The temperature was set and controlled at 298 K with a 535 l/h airflow rate in order to avoid any temperature fluctuations owing to sample heating during the magnetic field pulse gradients.
Amine or amino acid ester salts extraction runs reported in Table 2. Run 5. The 0.6 ml of D$_2$O solution contained 4.4 mg (0.029 mmol) of LiPF$_6$ and 4.6 mg (0.029 mmol) of (S)-phenylethylammonium hydrochloride (S)-14HCl was shaken for 1 min at r.t. with 0.8 ml of (S)-10 (2.8 mg, 0.0078 mmol) in CDCl$_3$ (0.01 M). The organic layer was dried (MgSO$_4$) and the spectrum taken. The runs 1-4 and 6-22 were carried by analogy with the corresponding host and guest.

Extraction, isolation, and rotation experiment of methyl phenylglycinate hexafluorophosphate

Host (aS,S)-11c (80 mg, 0.128 mmol) was dissolved in 0.7 ml of CDCl$_3$ to give a 0.18 M solution. This solution was used to extract 3 eq. of racemic methyl phenylglycinate hydrochloride rac-13·HCl (77.2 mg, 0.383 mmol) dissolved in 0.4 ml of an aqueous D$_2$O solution (0.92 M in guest) contained 0.0582 g (0.383 mmol) of LiPF$_6$. After equilibration at r.t. (about 30 min), the phases were carefully separated, and the meniscus was discarded. The organic phase was diluted with 1 ml of CH$_2$Cl$_2$ and extracted with three 0.8 ml portions of 0.1N HCl. The combined aqueous extracts were added to 2.6 ml of CH$_2$Cl$_2$ and aqueous ammonium hydroxide was added to adjust the pH to 10. The organic phase was withdrawn, the aqueous phase was reextracted with another 1.5 ml of CH$_2$Cl$_2$, and the combined organic extracts were dried with MgSO$_4$. The solvent was evaporated to give 10 mg of the amino ester 13 as an oil (0.06 mmol); [α]$_D^{20}$ = -40 (0.3, CHCl$_3$); [α]$_D^{20}$ = -33, [α]$_{578}^{20}$ = -35, [α]$_{546}^{20}$ = -39, [α]$_{436}^{20}$ = -76 (c 0.3, CH$_2$Cl$_2$), indicating an optical purity of 20-22 %, enriched in the (R)-enantiomer. (Cf. lit. [4] [α]$_{578}^{20}$ = -52.5, [α]$_{546}^{20}$ = -60.4, [α]$_{436}^{20}$ = -110.6 (c 2, CH$_2$Cl$_2$), for D-13, 32.6 % op). (Cf. lit. [27] for (S)-phenylglycine methyl ester: [α]$_D^{20}$ = 202.3 (0.49, CHCl$_3$)). $^1$H NMR δ: 1.88 (b.s, 2H), 3.77 (s, 3H), 4.65 (s, 1H), 7.28-7.38 (m, 5H). Examination of the NMR spectrum of this product showed the absence of 11c signals.

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