Synthesis of 5-substituted 7,9-dihydro-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-ones as anticonvulsant agents

Maria Zappalà,^a Nicola Micale,^{a,} Silvana Grasso,^{a,*} Frank S. Menniti,^b Giovambattista De Sarro,^c and Carlo De Micheli^d

 ^a Dipartimento Farmaco-Chimico, Università di Messina, viale Annunziata, 98168 Messina, Italy,
 ^b Pfizer Global Research and Development, Eastern Point Road, Groton, CT 06340, USA,
 ^c Dipartimento di Medicina Sperimentale e Clinica, Università di Catanzaro, Via T. Campanella, 88100 Catanzaro, Italy,
 ^d Istituto di Chimica Farmaceutica, Università di Milano, viale Abruzzi, 42, 20131 Milano, Italy E-mail: grasso@pharma.unime.it

Dedicated to Professor Vincenzo Tortorella on the occasion of his "Fuori Ruolo" status (received 19 Dec 03; accepted 05 Mar 04; published on the web 10 Mar 04)

Abstract

5-(4-Aminobenzyl)-7,9-dihydro-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**3**) and 7,9dihydro-5-[2-(pyridin-2-yl)-vinyl]-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**4**) were synthesized and screened as anticonvulsant agents in DBA/2 mice against sound-induced seizures. The new compounds are provided with anticonvulsant properties even if ED_{50} values are lower than those of prototype 5-(4-aminophenyl)-7,9-dihydro-8*H*-[1,3]dioxolo[4,5h][2,3]benzodiazepin-8-one (**2**) and of GYKI 52466 (**1**), a well-known noncompetitive AMPA receptor antagonist. Binding assays and functional tests were performed to evaluate the affinity for AMPA and kainate receptors.

Keywords: 7,9-Dihydro-8H-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-ones, anticonvulsant activity

Introduction

Ionotropic glutamate receptors (iGluRs) are the major excitatory neurotransmitter receptor proteins in the mammalian brain.^{1,2} As a class of membrane proteins, iGluRs are composed of subunits that span the membrane to form a small pore or channel, which is gated by glutamate, a neurotransmitter. When glutamate is released from a presynaptic neuron and binds to a postsynaptic glutamate receptor, the receptor rapidly changes its conformation and transiently forms an open ion channel, thus resulting in a change of the postsynaptic membrane potential. A postsynaptic potential of sufficient strength triggers an action potential, which will in turn

propagate the initial nerve impulse. The major function of iGluRs is to mediate fast synaptic neurotransmission underlying the basic activities of the brain, such as memory and learning. Overactivation of the receptors, on the other hand, has been implicated in a number of neurological diseases such as post-ischemia cell death, epilepsy, Huntington's chorea, and amyotrophic lateral sclerosis.¹ To date, three major subtypes of iGluRs have been identified: NMDA (*N*-methyl-D-aspartate), kainate, and AMPA (2-amino-3-(3-hydroxy-5-methylisoxazol-4-yl)propionic acid) receptors, and classified on the basis of amino acid sequences, pharmacological profiles, and biological functions.^{1,2}

5-(4-Aminophenyl)-8-methyl-9*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepine (**1**, GYKI 52466, Figure 1), a 2,3-benzodiazepine derivative, is the prototype of selective noncompetitive AMPA receptor antagonists acting *via* an allosteric site on the receptor complex.³ It possesses remarkable anticonvulsant properties^{4,5} and behaves as a neuroprotective agent in focal and global ischemia.⁶

Based on the anticonvulsant properties of the previously described^{7,8} 5-(4-aminophenyl)-7,9dihydro-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**2**), where the iminohydrazone portion of **1** was replaced by the iminohydrazide moiety, we designed 5-(4-aminobenzyl)-7,9-dihydro-8H-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**3**) and 7,9-dihydro-5-[2-(pyridin-2-yl)-vinyl]-8H-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**4**) (Figure 1) in order to explore the influence of the substituent appended at position 1 on the anticonvulsant activity. In particular we inserted in such a position the 4-aminobenzyl group, in order to interrupt the electronic connection between the aryl group and the unsaturated heterocyclic ring, and the 2-(pyridin-2-yl)-vinyl moiety which seems to play a role of utmost importance in a strictly-related class of new noncompetitive AMPA receptor antagonists.⁹

Herein, we report the synthesis and the anticonvulsant properties of derivatives **3** and **4** along with the results of binding assays and functional tests performed at both AMPA and kainate receptors.



Figure 1

Results and Discussion

Chemistry

The synthesis of novel compounds **3** and **4** was accomplished following the reaction sequence reported in Schemes 1-2.

Ketoester **6** was easily prepared *via* acylation of methyl 1,3-benzodioxol-5-ylacetate **5** with 4-nitrophenylacetic acid in the presence of excess phosphorous pentoxide. The subsequent treatment with hydrazine gave 7,9-dihydro-5-(4-nitrobenzyl)-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (**7**) in good yields. Reduction of the nitro group of **7**, carried out with Raney-Ni/ammonium formate, afforded target derivative **3**.



Scheme 1. (a) 4-nitrophenylacetic acid, P_2O_5 , $(CH_2Cl)_2$, rt, 16h; (b) NH_2NH_2 , MeOH, reflux, 48h; (c) Raney-Ni, ammonium formate, EtOH, reflux, 2h.

7,9-Dihydro-5-[2-(pyridin-2-yl)-vinyl]-8H-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (4) was easily synthesized by a Friedel-Craft acylation of 5 with acetic anhydride in the presence of tin(IV) chloride as the catalyst followed by a treatment with hydrazine to afford intermediate 9. Condensation of 9 with pyridine-2-carboxaldehyde to produce vinyl derivative 4 was conveniently accomplished with acetic anhydride and anhydrous zinc chloride in refluxing dioxane.

Physical and spectral data (¹H NMR) of the synthesized compounds are in agreement with the proposed structures. In compound 4, the stereochemistry around the C=C bond was assigned as *E*, on the basis of the coupling constant value (J = 16.2 Hz) between the two protons of the vinyl moiety.



Scheme 2. (a) acetic anhydride, SnCl₄, CH₂Cl₂, rt, 16h; (b) NH₂NH₂, MeOH, reflux, 48h; (c) pyridine-2-carboxaldehyde, acetic anhydride, ZnCl₂, dioxane, reflux, 16h.

Pharmacology

The anticonvulsant activity of derivatives **3** and **4** against audiogenic seizures was evaluated 30 min after intraperitoneal administration to DBA/2 mice, a strain genetically susceptible to sound-induced seizures. This test has been considered an excellent animal model for generalized epilepsy and for screening new anticonvulsant drugs.¹⁰ The results are compared with those previously reported for derivative **2** and reference compound **1** (Table 1).^{7,8}

As shown in Table 1, compound 4 possesses anticonvulsant properties lower than those of prototype 2 as well of lead compound 1 whereas derivative 3, even if unable to prevent the clonic phase of the audiogenic seizures, significantly reduces the tonic phase of the seizures; its ED_{50} value is comparable to that of GYKI 52466.

Table 1. Anticonvulsant activity of compounds **1-4** against audiogenic seizures in DBA/2 mice and TD_{50} values on locomotion assessed by rotarod test^a

Compds	ED ₅₀ , µmol/kg		TD ₅₀ , µmol/kg
	Clonic phase	Tonic phase	locomotor deficit
1	35.8 (24.4-52.4)	25.3 (16.0-40.0)	76.1 (47.5-122)
2	18.0 (10.0-32.5)	12.7 (6.13-26.2)	101 (52.0-194)
3	>80	24.1 (13.6-42.7)	> 150
4	81.2 (52.2-126)	65.5 (46.7-91.8)	> 150

^a All compounds were given ip (at doses spanning the range 3.3-200 μ mol/kg) 30 min before auditory stimulation. All data were calculated according to the method of Litchfield and Wilcoxon;¹⁷ 95% confidence limits are given in brackets. At least 32 animals were used to calculate each ED₅₀ and TD₅₀ value.

For this reason, compound **3** was also examined for its ability to displace [³H]CP-526,427 from the corresponding binding site of the AMPA receptor complex (Table 2). The inhibition of [³H]CP-526,427 specific binding (3 nM) to rat forebrain membranes was evaluated as previously described.^{11,12} In this assay, the quinazolinone CP-465,022 was used as a positive control; its IC₅₀ values spanned the range 20-40 nM. Compound **3** turned out to be totally inactive (IC₅₀ >100 μ M), differently to that observed¹³ for compound **2** which showed IC₅₀ = 32 μ M, a value similar to that reported for GYKI 52466 (IC₅₀ = 12.6 μ M).

Furthermore, compound **3** was tested for its ability to inhibit the kainate-induced increase of the $[Ca^{2+}]i$ in a primary culture of rat cerebellar granule cells (CGC) which express AMPA receptors (Table 2); GYKI 52466 was used as the control. The same test was carried out in rat HEK293 cells expressing GluR5, a kainate receptor subtype, stimulated by domoic acid; SYM 2081 was used as the control.

The results of the CGC test confirmed the data of the binding experiments, namely compound **3** at 100 μ M produced solely a 20% inhibition of calcium influx, whereas derivative **2**

turned out to be slightly more potent than 1 (IC₅₀ 13 μ M for 2 vs 22 μ M for 1) (Table 2). In the iGluR5/HEK293 cells test, compound 3, assayed up to 10 μ M, showed neither agonistic nor antagonistic activity.

Table 2. Pharmacological data of compounds 2-3. and of reference compounds GYKI 52466 and

 SYM 2081

Compd	[³ H]CP-526,427 ^a	$KA-[Ca^{2+}]i$	Domoic acid-[Ca ²⁺]i
I	$IC_{50} (\mu M)$	IC_{50} (μ M) or 1%	$IC_{50} (\mu M)$
2	32	13	-
3	>100	20%	>10
GYKI 52466	12.6	22	-
SYM 2081	-	-	0.047

^a Applied (3 nM) to rat forebrain membranes.

To sum up, the results reported in this study suggest that the 4-aminophenyl group directly linked to the unsaturated heterocycle is a fragmental requirement necessary to confer anticonvulsant activity to compounds structurally related to derivative 2. The insertion of a methylene between the aryl group and the unsaturated heterocyclic ring to interrupt their electronic connection or an extension of their conjugation through the insertion of a vinyl moiety, in both cases, bring about a drastic reduction in the anticonvulsant activity.

Experimental Section

General Procedures. Melting points were determined on a Kofler hot stage apparatus and are uncorrected. Elemental analyses were carried out on a C. Erba Model 1106 Elemental Analyzer for C, H and N) and the results are within $\pm 0.4\%$ of the theoretical values. Merck silica gel 60 F₂₅₄ plates were used as analytical TLC; column chromatography was performed on Merck silica gel 60 (70-230 mesh). IR spectra were obtained on a Perkin Elmer Spectrum BX FT-IR as nujol mulls. ¹H-NMR spectra were recorded in CDCl₃ by means of a Varian Gemini 300 spectrometer. Chemical shifts are expressed in δ (ppm) relative to TMS as internal standard and coupling constants (*J*) in Hz.

Methyl 6-[(4-nitrophenyl)acetyl]-1,3-benzodioxol-5-ylacetate (6). 4-Nitropheylacetic acid (606 mg, 3.3 mmol) and phosphorous pentoxide (2.0 g) were added to a stirred 1,2-dichloroethane solution (50 mL) of methyl 1,3-benzodioxol-5-ylacetate 5^{14} (500 mg, 2.6 mmol). The mixture was further stirred at room temperature overnight, then water (30 mL) was cautiously added and the mixture extracted with chloroform (2 x 30 mL). The organic layer was separated and sequentially treated with 10% NaOH (30 mL), brine (30 mL) and water (2 x 30 mL). The organic phase was dried (Na₂SO₄) and the solvent removed under reduced pressure

to yield crude **6** which was purified by crystallization with diethyl ether/light petroleum. Mp 128-131°C (orange powder) (745 mg, 81%) $R_f = 0,36$ (Et₂O/light petroleum 60/40); IR (nujol): 1727, 1665 cm⁻¹. ¹H NMR (CDCl₃): 3.66 (s, 3H, CH₃), 3.85 (s, 2H, CH₂-1), 4.30 (s, 2H, COCH₂Ar), 6.07 (s, 2H, OCH₂O), 6.74 (s, 1H, H-6), 7.33 (s, 1H, H-3), 7.40 (d, 2H, *J* = 8.8, H-2',6'), 8.20 (d, 2H, *J* = 8.8, H-3',5'). Anal. calcd. for C₁₈H₁₅NO₇: C, 60.51; H, 4.23; N, 3.92. Found: C, 60.36; H, 4.17; N, 4.04.

7,9-Dihydro-5-(4-nitrobenzyl)-8*H***-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (7).** Hydrazine hydrate (0.28 mL, 5.82 mmol) was added to a solution of compound **6** (650 mg, 1.82 mmol) in methanol (40 mL); the mixture was acidified to pH 3 by addition of 3N HCl then heated at reflux for 48 h. The solvents were removed under reduced pressure and the residue was dissolved with EtOAc and poured into water. The organic layer, dried over Na₂SO₄, was evaporated under reduced pressure and the residue was dissolved with EtOAc/cyclohexane, 60:40) to afford pure **7**. Mp 229-232 °C (beige powder) (424 mg, 60%) R_f = 0,39 (EtOAc/cyclohexane 60/40); IR (nujol): 3179, 1684 cm⁻¹. ¹H NMR (CDCl₃): 3.22 (s, 2H, CH₂-5), 4.19 (s, 2H, CH₂-1), 6.04 (s, 2H, OCH₂O), 6.70 (s, 1H, H-9), 6.94 (s, 1H, H-6), 7.38 (d, 2H, J = 8.8, H-2',6'), 8.13 (d, 2H, J = 8.8, H-3',5'), 8.37 (bs, 1H, NH). Anal. calcd. for: C₁₇H₁₃N₃O₅: C, 60.18; H, 3.86; N, 12.38. Found: C, 60.32; H, 3.67; N, 12.14.

5-(4-Aminobenzyl)-7,9-dihydro-8*H***-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (3).** A suspension of **7** (392 mg, 1.15 mmol) and Raney-Ni (60 mg) in EtOH (40 mL) was stirred with ammonium formate (362 mg, 5.75 mmol). The mixture was refluxed for 2 h and then filtered on celite. The organic layer was evaporated under reduced pressure and the residue, dissolved in CHCl₃, was washed with saturated NaCl to remove ammonium formate. The organic layer, dried over Na₂SO₄, was evaporated under reduced pressure and the residue by a silica gel column chromatography eluting with EtOAc. Mp 104-107 °C (pale beige powder) (317 mg, 82%) R_f = 0,42 (EtOAc); IR (nujol): 3350, 3220, 1667 cm⁻¹. ¹H NMR (CDCl₃): 3.20 (s, 2H, CH₂-5), 3.61 (bs, 2H, NH₂), 3.96 (s, 2H, CH₂-1), 6.00 (s, 2H, OCH₂O), 6.57 (d, 2H, *J* = 8.5, H-3',5'), 6.67 (s, 1H, H-9), 6.96 (d, 2H, *J* = 8.5, H-2',6'), 6.97 (s, 1H, H-6), 8.35 (bs, 1H, NH). Anal. calcd. for C₁₇H₁₅N₃O₃: C, 66.01; H, 4.89; N, 13.58. Found: C, 66.23; H, 4.76; N, 13.33.

Methyl 6-acetylbenzo[1,3]dioxol-5-ylacetate (8). Tin(IV) chloride (0.1 M 8 mL, 8 mmol) and acetic anhydride (0,63 mL, 6.70 mmol) in CH₂Cl₂ (20 mL) were added to a cooled (0-5°C) and stirred solution of 3,4-methylenedioxyphenyl acetic acid methyl ester 5 (1 g, 5.15 mmol) in 20 ml of the same solvent. The ice-bath was removed and the mixture was stirred at 20°C overnight, then poured into water and the product was isolated in ether and dried over Na₂SO₄. The solvent was removed under reduced pressure and the residue purified by treatment with diethyl ether. Mp = 111-114 °C (beige powder) (1.14 g, 94%) $R_f = 0.52$ (Et₂O/light petroleum 60/40); IR (nujol): 1719, 1659 cm⁻¹. ¹H NMR (CDCl₃): 2.53 (s, 3H, COCH₃), 3.71 (s, 3H, COOCH₃), 3.85 (s, 2H, CH₂-1), 6.05 (s, 2H, OCH₂O), 6.72 (s, 1H, H-6), 7.31 (s 1H, H-3), Anal. calcd. for C₁₂H₁₂O₅: C, 61.02; H, 5.12. Found: C, 61.22; H, 5.01.

7,9-Dihydro-5-methyl-8*H***-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one** (**9**). Compound **9** was synthesized with a procedure similar to that reported for derivative **7** starting from compound **8**

(1.14 g, 4.83 mmol), and purified by a silica gel column chromatography eluting with EtOAc/MeOH (90/10). Mp 251-253°C (white powder) (748 mg, 72%) $R_f = 0.79$ (EtOAc/MeOH 90/10); IR (nujol): 3185, 1678 cm⁻¹. ¹H NMR (CDCl₃): 2.44 (s, 3H, CH₃), 3.34 (s, 2H, CH₂-5), 6.03 (s, 2H, OCH₂O), 6.74 (s, 1H, H-9), 6.92 (s, 1H, H-6), 8.20 (bs, 1H, NH). Anal. calcd. for C₁₁H₁₀N₂O₃: C, 60.55; H, 4.62; N, 12.84: Found: C, 60,37; H, 4.86; N, 12.62.

E-7,9-Dihydro-5-[2-(pyridin-2-yl)-vinyl]-8*H*-[1,3]dioxolo[4,5-h][2,3]benzodiazepin-8-one (4). An anhydrous dioxane solution (40 ml) of 9 (748 mg, 3.43 mmol) was treated with 2pyridinecarboxaldehyde (0.33 ml, 3.43 mmol), acetic anhydride (2 mL) and an excess of anhydrous ZnCl₂. The reaction mixture was refluxed overnight, then poured into water and extracted with chloroform (2 x 50 mL). The organic layer, dried over Na₂SO₄, was evaporated under reduced pressure and the residue was purified by a silica gel column chromatography eluting with CCl₄/*i*-PrOH (95/5). Mp 240-243°C (white needles) (643 mg, 61%) R_f = 0.24 (CCl₄/*i*-PrOH 95/5); IR (nujol): 3188, 1676 cm⁻¹. ¹H NMR (CDCl₃): 3.39 (s, 2H, CH₂-5), 6.06 (s, 2H, OCH₂O), 6.82 (s, 1H, H-9), 7.03 (s, 1H, H-6), 7.04 (d, 1H, *J*_{trans} = 16.2, CH_{vinyl}) 7.20-8.67 (m, 4H, Py-H), 7.59 (d, 1H, *J*_{trans} = 16.2, CH_{vinyl}), 8.57 (bs, 1H, NH). Anal. calcd. for C₁₇H₁₃N₃O₃: C, 66.44; H, 4.26; N, 13.67. Found: C, 66.31; H, 4.37; N, 13.78.

Audiogenic seizures test in DBA/2 mice.¹⁵ DBA/2 mice (8-12 g, 22-25 days old) were purchased from Charles River (Calco, Como, Italy). Groups of 10 mice of either sex were exposed to auditory stimulation 30 min following administration of vehicle or each dose of drugs studied. The compounds were given ip (0.1 mL/10 g of body weight of the mouse) as a freshly-prepared solution in 50% DMSO and 50% sterile saline (0.9% NaCl). Individual mice were placed under a hemispheric perspex dome (diameter 58 cm) and 60 s were allowed for habituation and assessment of locomotor activity. Auditory stimulation (12-16 kHz, 109 dB) was applied for 60 secs or until tonic extension occurred and induced a sequential seizure response in control DBA/2 mice, consisting of an early wild running phase, followed by generalized myoclonus and tonic flexion and extension, sometimes followed by respiratory arrest. The control and drug-treated mice were scored for latency to and incidence of the different phases of the seizures.¹⁶

Statistical analysis. The ED_{50} values of each phase of the audiogenic seizure were determined for each dose of compound administered, and dose-response curves were fitted using a computer program by the method of Litchfield and Wilcoxon.¹⁷ The median toxic dose (TD₅₀) values were estimated using the method of Litchfield and Wilcoxon.¹⁷

Acknowledgments

This work was financially supported by the Ministero dell'Istruzione, dell'Università e della Ricerca (MIUR - COFIN2003) - Rome, Italy.

References

- 1. Dingledine, R.; Borges, K.; Bowie, D.; Traynelis, S. F. Pharmacol. Rev. 1999, 51, 7.
- 2. Hollmann, M.; Heinemann, S. Annu. Rev. Neurosci. 1994, 17, 31.
- 3. Sólyom, S.; Tarnawa, I. Curr. Pharm. Design 2002, 8, 913 and references cited therein.
- 4. Donevan, S. D.; Yamaguchi, S.; Rogawski, M. A. J. Pharmacol. Exp. Ther. 1994, 271, 25.
- 5. Chapman, A. G.; Smith, S. E.; Meldrum, B. S. Epilepsy Res. 1991, 9, 92.
- 6. Smith, S. E.; Meldrum, B. S. Stroke 1992, 23, 861.
- 7. De Sarro, A.; De Sarro, G.; Gitto, R.; Grasso, S.; Micale, N.; Quartarone, S.; Zappalà M. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 971.
- 8. Micale, N.; Zappalà, M.; Grasso, S. Il Farmaco 2003, 58, 351.
- Welch, W. M.; Ewing, F. E.; Huang, J.; Menniti, F. S.; Pagnozzi, M. J.; Kelly, K.; Seymour, P. A.; Guanowsky, V.; Guhan, S.; Guinn, M. R.; Critchett, D.; Lazzaro, J.; Ganong, A. H.; DeVries, K. M.; Staigers, T. L.; Chenard, B. L. *Bioorg. Med. Chem. Lett.* 2001, 11, 177.
- (a) Chapman, A. G.; Croucher, M. J.; Meldrum, B. S. Arzneim. Forsch. 1984, 34, 1261. (b) Engstrom, F. L.; Woodbury, D. M. Epilepsia 1988, 29, 389.
- Menniti, F. S.; Chenard, M. B.; Collins, M. F.; Ducat, M. F.; Elliot, M. L.; Ewing, F. E.; Huang, J. I.; Kelly, K. A.; Lazzaro, J. T.; Pagnozzi, M. J.; Weeks, J. L.; Welch, W. M.; White, W. F. *Mol. Pharmacol.* **2000**, *58*, 1310.
- 12. Parks, T. N.; Artman, L. D.; Alasti, N.; Nemeth, E. F. Brain Res. 1991, 552, 13.
- 13. Grasso, S.; Micale, N.; Zappala, M.; Galli, A.; Costagli, C.; Menniti, F. S.; De Micheli, C. *Bioorg. Med. Chem. Lett.* **2003**, *13*, 443.
- 14. Cabedo, N.; Andreu, I.; Ramirez de Arellano, M. C.; Chagraoui, A.; Serrano, A.; Bermejo, A.; Protais, P.; Cortes, D. J. Med. Chem. 2001, 44, 1794.
- 15. Collins, R. L. In *Experimental Models of Epilepsy;* Purpura, P.; Penry, J. K.; Tower, D.; Woodbury, D. M.; Walter, R. Eds; Raven Press: New York, 1972, pp 347-372.
- 16. De Sarro, G. B.; Croucher, M. J.; Meldrum, B. S. Neuropharmacology 1984, 23, 526.
- 17. Litchfield, J.; Wilcoxon, F. J. Pharmacol. Exp. Ther. 1949, 96, 99.